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Preliminary study of urinary biomarker, miR-21, for kidney injury detection

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Abstract: Herbicide, namely glyphosate, has long been reported to cause adverse effects in human 11 especially the induction of kidney injury and disturbance of kidney functions. Several biomarkers 12 were demonstrated to be used for early kidney injury detection. However, some biomarkers have a 13 limitation and low sensitivity to detect early phase of kidney injury. Therefore, we aimed to estab-14 lish two urinary kidney injury biomarkers, urinary kidney injury molecule-1 (KIM-1) and mi-15 croRNA-21 (miR-21), in glyphosate-exposed farmers. Two spot urine samples consisting of pre-16 work urine sample (before glyphosate application) and post-work urine sample (after glyphosate 17 application 24 hours) were obtained. Urinary KIM-1 concentrations and miR-21 were analyzed. The 18 average level of urinary KIM-1 was 1.34 µg/g Cr in pre-work urine and 1.26 µg/g Cr in post-work 19 urine. The average fold of miR-21 expression was 1.40 and 1.21-fold in the pre- and post-work urine 20 sample. There were no significant differences of the two biomarkers between the pre-work and post-21 work urine sample. However, the Δ miR-21 expression moderately correlated with the volume of 22 glyphosate used. Although two biomarkers were not significant different between pre- and post-23 work urine sample, the miR-21 expression may relate with the dose of glyphosate application in 24 farmers. The number of subjects in this is limited, therefore a population size should be increased 25 in future study. 26

Keywords: glyphosate, urinary biomarker, KIM-1, miR-21, occupational exposure, kidney injury

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1. Introduction

Glyphosate is globally used in agricultural activities to control weeds in cropped 30 and non-cropped fields. The glyphosate and glyphosate-based herbicides toxicity has 31 long been reported to exert harmless in human and animal [1]. Therefore, the potential 32 health risks from glyphosate and glyphosate-based herbicides exposure have long been 33 concerned. The farmers are faced with increasing risks related to expose glyphosate dur-34 ing working in farm through inhalation, skin absorption, and oral ingestion. After that, 35 glyphosate and its metabolites were excreted through kidney [1]. According to the study 36 of Gao et al., 2019, the renal proximal tubule was identified as a main target of glypho-37 sate and glyphosate-based herbicides [2]. Also, several studies reported that the history 38 exposure of glyphosate associated with changes in kidney injury and functions [3-7]. 39

Many biomarkers such as creatinine, blood urea nitrogen, and urine markers of kidney injury (epithelial cells, tubular casts, urinary concentrating ability, etc.) were generally used to evaluate kidney injury and function [8]. However, some biomarkers such 42

BMB Conference 2021.

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as serum creatinine (sCr) and urine output could be detected in late kidney injury and 43 showed low sensitivity and specificity [9]. In addition, urinary kidney injury molecule-1 44 (KIM-1), urinary beta2-microglobulin (B2M), urinary total protein, urinary albumin, 45 urinary clusterin, urinary cystatin c (Cys C), and urinary trefoil factor 3 (TFF-3) have 46 been reported to show high sensitivity to detect early acute kidney injury (AKI) [9-11]. 47 In the case of KIM-1, it was highly localized in proximal tubule cells and can be detected 48 in urine [12]. This biomarker has been reported to use for early AKI diagnosis and xeno-49 biotic-induced kidney injury monitoring [13, 14]. The meta-analysis demonstrated that 50 urinary KIM-1 level was specific to diagnose AKI with specificity at 86.0% and sensitiv-51

glomerular filtration rate (eGFR) [16]. With the recent developments in high-throughput technologies, the pace of advances in the functional genomics and proteomics that are applicable to establish novel biomarker for several diseases especially kidney disease has noticeably accelerated [17]. The microRNAs (miRNAs) have been recently emerged as promising biomarker candidates for several diseases such as cancer, Alzheimer's, and kidney diseases [18-20]. The miRNAs are endogenous short RNA molecules and play important role to regulate mRNA expression by mRNA destabilization and translational repression [21]. The great advantage of miRNAs to serve potentially excellent biomarker was the stability in biological samples such as blood, serum, urine, and other sources [22]. The miR-21 function has been proposed to be involved in cellular apoptosis, inflammation, and fibrosis signaling pathways in AKI [18, 23, 24]. In the model of drug-induced nephrotoxicity, the upregulation of miR-21 was found in damaged kidney in order to decrease lipid peroxidation and reactive oxygen species as well as deregulate metabolic processes [25]. Previous study illustrated that the significant alteration of miR-21 was observed in both acute as well as chronic animal models of kidney injury [26]. Moreover, the regulation of TGF- β signaling which directly acted to control reactive oxygen species (ROS) production was reported to implicate with miR-21in tubulointerstitial fibrosis associating with kidney dysfunction [27]. Hence, it was likely that the function of miR-21 in kidney disease was involved to response ROS generation.

ity at 74.0% [15]. In addition, the KIM-1 level was represented to predict kidney function

deterioration since this level was significantly correlated with a decrease in estimated

Taken together, the proposed mechanism of herbicide namely glyphosate on kidney injury was based on ATP depletion via uncoupling of oxidative phosphorylation, cytochrome C activation, and macromolecular oxidation resulting to induce kidney cell death [28]. Therefore, we assumed that ROS will be generated in kidney cells after glyphosate exposure. Then, the kidney cell biomarker such as KIM-1 and miR-21 will be released from injured kidney cells and could be initially detected in the urine. The objective of this study is to evaluate the KIM-1 level and miR-21 expression to establish a novel biomarker for early kidney injury detection among glyphosate-exposed farmers.

2. Materials and Methods

2.1 Population, questionnaire, and urine collection

This study was conducted in Long District, Phrae Province, Thailand. Ninety-six84farmers who practically applied glyphosate herbicide were recruited into our study to85conduct face-to-face interview and collect urine. The questionnaire consisted of demo-86graphic data, personal and health history, work characteristics (number of years working87with herbicides, time to exposure glyphosate, type of work, volume, concentration, and88frequency), and personal protective equipment (PPE) use. All full urinary voided spot89sample were collected over the exposure assessment period (before herbicide application)90

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2.2 Quantification of urinary glyphosate	92
The urinary glyphosate was determined by a liquid chromatography-tandem mass $C_{\rm exp}$	93 04
spectrometry (LC–MS/MS) according to previous publication [29]. The farmer's urine	94
which had urinary glyphosate concentration over limit of quantification (LOQ) by 5	95 06
μ g/L in period of exposure were further selected to determine kidney injury biomarkers.	96
Two spot urine samples were chosen consisting of pre-work urine sample (before	97
glyphosate application) and post-work urine sample (after glyphosate application 24	98
hours).	99
2.3 Determination of urinary creatinine	100
To normalized urine volume in kidney biomarker determination, urinary creatinine	100
was quantified by ARCHITECT [™] clinical chemistry analyzer (Abbott, USA) at Maharaj	101
Nakorn Chiang Mai Hospital Central Laboratory, Faculty of Medicine, Chiang Mai Uni-	102
versity. The urinary creatinine was expressed as mg/dL.	103
versity. The armany eleatinine was expressed as mg/a2.	101
2.4 Determination of Human Kidney injury molecule 1 (KIM-1)	105
To determine the kidney injury marker in glyphosate-exposed farmers, the KIM-1	106
level in pre- and post-work urine samples was measured by ELISA according to the	107
manufacturer's instructions (CUSABIO, China). One-hundred microliters of urine sam-	108
ple were added into human antibody-coated ELISA plates and the plate was then incu-	109
bated at 37 °C for 2 hours. Next, the biotin-conjugated antibody specific for KIM-1 was	110
added into the plate. After incubation for 1 hour, the reaction color was developed by	111
the chemical reaction between the tetramethylbenzidine (TMB) reagent and streptavidin	112
conjugated with horseradish peroxidase (HRP). The absorbance was measured at a	113
wavelength of 450 nm using a microplate reader (Synergy™ H4, BioTek Instruments,	114
Inc., USA). The concentration of urinary KIM-1 was calculated to compare with standard	115
curve and normalized with urinary creatinine. The urinary KIM-1 level was expressed as	116
μg/g creatinine.	117
2.5 MicroRNA-21 expression	118
To evaluate the expression of urinary miR-21 in glyphosate-exposed farmers, the	119

To evaluate the expression of urinary miR-21 in glyphosate-exposed farmers, the miR-21 expression in pre- and post-work urine samples was quantified by quantitative real-time polymerase chain reaction (qRT-PCR). First, the urine was centrifuged by ultra-centrifugation at 200,000 g 4 °C for 1 hour to harvest urinary exosome. Then, the miR-21 in urinary exosome was extracted by Nucleozol reagent (Toyobo, Japan) according to the manufacturer's protocol. Total urinary miRNA was reverse transcribed into cDNA by the Mir-XTM miRNA First-Strand Synthesis Kit (Takara Bio Company, USA). The expression of urinary miR-21 was determined by commercial Mir-X miRNA qRT-PCR TB Green Kit (Takara Bio Company, USA). The primer for miR-21 was demonstrated in Table 1. In brief, the PCR parameters for gene amplification were 40 cycles at 95 °C for 5 min for the denaturation, at 95 °C for 12 s for annealing, and at 65 °C for 50 s for extension. The expression of the target miR-21 level was analyzed by the $2^{-\Delta\DeltaCt}$ method using U6 snRNA

as a control according to the manufacturer's protocol. The miR-21 expression was ex-

Table 1. The sequences of miRNA-21 primers used for qRT-PCR.

Gene target	Sequence	Reference
miR-21-5p	5'-CGGCGGTAGCTTATCAGACTGA	[30]

2.6 Statistical analysis methods

pressed as fold.

Statistical analysis was performed using the SPSS for Windows, Version 16.0. (Chi-135cago, SPSS Inc; 2007) and the GraphPad Prism version 8.3.0 for windows (GraphPad Soft-136ware, San Diego, California USA, www.graphpad.com). All demographic data (gender,137age, number of years working with herbicides, time to exposure glyphosate, type of138

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work, volume, concentration of glyphosate in urine, and personal protective equipment139usage) were described and analyzed by descriptive statistics. The levels of urinary KIM-1140and miR-21 expression in pre-work and post-work urine samples were expressed as me-141dian (minimum – maximum) and compared using the Wilcoxon signed-rank test. Spear-142man's rank correlation test was used to evaluate a relationship between urinary bi-143omarkers and demographic data.144

3. Results

The urinary glyphosate level was analyzed in 96 participants. The results showed 146 that the urinary glyphosate level which was greater than limit of quantification (LOQ) by 147 $5 \mu g/L$ were observed in 17 participants. Therefore, these participants were selected for 148 urinary biomarker quantification. The demographic characteristics were represented in 149 Table 2. Most participants were male (64.7%) and aged between 37 - 60 years old. 150 The average of farming experience of all subjected was 28 ± 13 years and the average 151 exposure time to glyphosate was approximately 5 hour per day. All of participants had 152 the responsibility to spray glyphosate on the agricultural area. Regarding behavior in 153 using PPE, participants usually wore gloves (52.9%), masks (47.1%), boots (100%), and 154 long-sleeved shirts (100%). The concentration of urinary glyphosate was 37.9 ± 37.5 (10.26) 155 -170.43) µg/g creatinine. 156

Table 2. Demographic data of the subjects

Characteristics	N (%) / mean ± SD
Gender	
Male	11 (64.7)
Female	6 (35.3)
Years working with herbicides (years)	28 ± 13
Time to exposure glyphosate (hours)	5 ± 2.8
Type of work	
Mixing and loading glyphosate	13 (76.5)
Spraying pesticides	17 (100)
Cleaning and collecting equipment	14 (82.4)
Use of Personal Protective Equipment (PPE)	
Mask	8 (47.1)
Gloves	9 (52.9)
Boots	17 (100)
Long-sleeved shirt	17 (100)
Concentration of glyphosate (µg/g creatinine)	37.9 ± 37.5

The comparison of urinary KIM-1 level and miR-21 expression between pre- and post-working urine samples are shown in Figure 1. The average urinary KIM-1 level in the pre-working and post-working urine samples were $1.34 (0.60 - 17.82 \,\mu\text{g/g Cr})$ and $1.26 (0.26 - 9.69 \,\mu\text{g/g Cr})$, respectively. However, no statistical significance was observed between pre- and post-working urine samples. The expression of urinary miR-21 was compared between pre- and post-working urine samples. The average miR-21 expression was 1.4 (0.07 - 12.97) fold in the pre-working urine sample and 1.21 (0.04 - 7.67) fold in the post-working urine sample. There were no significant differences of the urinary miR-21 expression between the pre-working and post-working urine sample.

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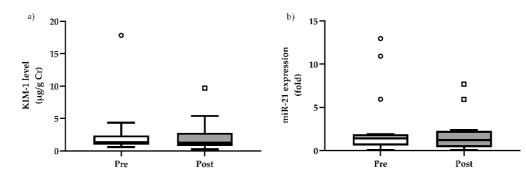


Figure 1. Two urinary biomarkers for kidney injury detection were investigated in glyphosate-exposed farmers. (a) The urinary KIM-1 level and (b) miR-21 expression of glyphosateexposed workers in the pre-work (open bar) and post-work (solid bar) urine sample. The data are represented as median (minimum – maximum).

To evaluate the correlation between kidney injury markers and demographic data, 174 the Δ KIM-1 level and Δ miR-21 expression were calculated to represent the changes of biomarker during glyphosate application. The biomarker level in pre-working urine sample was subtracted from the post-working urine sample to generate Δ KIM-1 level and Δ miR-21 expression. The result showed that Δ miR-21 expression was to be a moderate positive correlation with volume of glyphosate (r = 0.499). 177

	∆miR-21	Age	Years to	Exposure	Volume of
	Expression	(years)	working with	assessment	glyphosate
			pesticides	period	use (L)
				(hours)	
ΔKIM-1	0.471	0.409	0.478	0.104	0.259
Level					
ΔmiR-21	-	0.345	0.362	0.185	0.499*
Expression					

Table 3. Spearman's correlation (r) between urinary biomarkers and demographic data *p < 0.05

4. Discussion

Many studies have been reported the association between glyphosate exposure and 182 kidney injury [1, 31, 32]. The quantification of urinary biomarkers was performed in our 183 study to establish suitable biomarker for early kidney injury monitoring in glyphosate-184 exposed farmers. The amount of urinary KIM-1 level and miR-21 expression was not sig-185 nificant different between pre-work and post-work urine samples of farmers. Previous 186 study demonstrated that the significant level of urinary KIM-1 was found in herbicide-187 exposed farmers. De Silva et al., 2016 showed that urinary KIM-1 level in agricultural 188 workers living in chronic kidney disease endemic areas was significantly higher than this 189 level in control subjects from the same locations. The urinary KIM-1 was suggested to be 190 early kidney damage markers for identify CKD suspected cases [33]. However, the uri-191 nary KIM-1 level was not significantly increased in patients with AKI causing from acute 192 PQ poisoning [34]. In addition, Abdul et al., 2021 demonstrated that the urinary glypho-193 sate or paraquat level did not correlate with the urinary KIM-1 level [32]. 194

The study of miR-21 expression was performed to identify cancer, kidney, and cardiovascular diseases [35, 36]. Regarding kidney disease, the urinary KIM-1, miR-21, miR-200c, and miR-423 in both acetaminophen overdose patients with AKI and without AKI 197

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diagnosis were significantly higher than those level in healthy controls [37]. Although the 198 study of urinary miR-21 expression in herbicide-exposed workers have been less 199 reported, many publications have been focused the response of other microRNAs in pes-200 ticide intoxication. The expression of miR-223, miR-518d-3p, miR-517b, miR-597, and miR-201 28-5p were found to be significant increase in farmer during post-harvest seasons of agri-202 cultural activity. Moreover, the positive dose-response trend of the miRNAs in the class 203 of miR-223, miR-518d-3p, miR-597, miR-517b, and miR-133b with total dialkylphosphates 204 (organophosphate insecticide metabolites) were remarkably observed [23]. Also, the ex-205 pression of miRNAs (miR-29a-3p, miR-23b-3p, miR-19b-3p, miR-130a-3p, miR-125a-5p, 206 and miR-106b-5p) were significantly up-regulated in paraquat-treated lung cell lines. 207 These up-regulated miRNAs mediated the protein insertion into mitochondrial mem-208 brane involved in apoptotic signaling pathway [38]. Our finding demonstrated the signif-209 icant positive correlation between Δ miR-21 expression and volume of glyphosate use (L). 210 It was the first to report the relationship of miR-21 biomarker with the glyphosate usage. 211 It was assumed that high volume of glyphosate usage in farms was key parameter demon-212 strating the high risk of intake exposure. The high dose of glyphosate could induce kidney 213 injury as described above. Subsequently miR-21 was early detected and could be used for 214 kidney injury prediction. However, many factors probably influenced on urinary bi-215 omarker levels in our study leading to insignificant results between pre- and post-urine 216 samples. These could be explained with following reasons. First, the exposure dose during 217 work-task was relatively low in comparison with the previous study of Abdul et al., 2021. 218 They found the significant of renal biomarkers such as serum cystatin C and neutrophil 219 gelatinase-associated lipocalin (NGAL) in glyphosate users who showed the urinary 220 glyphosate level between 33.1 – 827.3 μ g/g Cr (median 224.3 μ g/g Cr) which was higher 221 than glyphosate level in our study [32]. Moreover, all farmers used PPE for instance mask, 222 gloves, boots, and long-sleeved shirt to reduce the risk of exposure [31, 39, 40]. 223

The limitation of this study was no biomarker data of a healthy control group (normal population), who do not apply herbicides. The sample size for biomarker quantification is limited. Although two biomarkers were not significant between pre- and post-work urine sample, it might be concluded that the miR-21 expression related with the dose of glyphosate application in farmers. 228

5. Conclusions

In this work, there was no significant differences of selected biomarkers between prework and post-work. The Δ miR-21 expression positively correlated with volume of glyphosate used with the spearman's rank correlation analysis. The miR-21 could be used as a marker to predict early renal injury in occupational exposure. However, more study needs to be done. 230 231 232 233 234

Supplementary Materials: None

Author Contributions: KW and SK conceived the present idea. KW, SK and RS were involved in
planning and supervised the work. KW, SK, KK and UI contributed to sample collection and man-
agement. KK and UI contributed to interviewing of the participants. KK and SK carried out the
experiments. KK, SK, and KW contributed to the interpretation of the results and data analysis. KK,
SK, and KW contributed to original draft preparation. All authors have read and agreed to the pub-
lished version of the manuscript.236
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Funding: This work was supported by Faculty of Medicine, Chiang Mai University, Grant No. 107-2422563 and 108-2563. The funding body had no role in the design and execution of this study or inter-243pretation of the data.244

Institutional Review Board Statement: This study was approved by the Ethics Committee of the245Faculty of Medicine, Chiang Mai University, Thailand (Study code: FOR-2562-06349/Research ID:2466349 and Study code: FOR-2563-06996/Research ID: 6996).247

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	Informed Consent Statement: Informed consent was obtained from all subjects involved in the study.	248 249
	Data Availability Statement: None	250
	Acknowledgments: The authors would like to express great appreciation to all farmworkers from Thung Lang Subdistrict, Long District, Phrae province. All help and supports from Pha-Chap health promoting hospital during this study.	
	Conflicts of Interest: The authors declare no conflict of interest.	254
References		255
1. Peillex, C.; Pelletier, M., Immunotoxicol 2020, 17 (1), 163-	The impact and toxicity of glyphosate and glyphosate-based herbicides on health and immunity.	J 256 257
 Gao, H.; Chen, J.; Ding tor is involved in glyphosate-ind Wunnapuk, K.; Gobe, G 	g, F.; Chou, X.; Zhang, X.; Wan, Y.; Hu, J.; Wu, Q., Activation of the N-methyl-d-aspartate recept duced renal proximal tubule cell apoptosis. Journal of Applied Toxicology 2019, 39 (8), 1096-1107. G.; Endre, Z.; Peake, P.; Grice, J. E.; Roberts, M. S.; Buckley, N. A.; Liu, X., Use of a glyphosate otoxicity model to investigate a panel of kidney injury biomarkers. Toxicology letters 2014, 225 (1)	258 259 2- 260
4. Indirakshi, J.; Sunnesh, Ram, R.; Kumar, V. S., Toxic epi	A.; Aruna, M.; Reddy, M. H. K.; Kumar, A. C.; Chandra, V. S.; Sangeetha, B.; Katyarmal, D. dermal necrolysis and acute kidney injury due to glyphosate ingestion. Indian journal of critical care al publication of Indian Society of Critical Care Medicine 2017, 21 (3), 167.	.; 263
	Y.; Hirano, K., Renal cortical hypoperfusion caused by glyphosate-surfactant herbicide. Clinical and	
6. Zhang, F.; Xu, Y.; Liu, Z	X.; Pan, L.; Ding, E.; Dou, J.; Zhu, B., Concentration Distribution and Analysis of Urinary Glypho pationally Exposed Workers in Eastern China. International Journal of Environmental Research and	- 268
and occupational exposure to H Health 2015, 14 (1), 6.	gama, P.; Agampodi, S.; Wijewardane, C.; Gunatilake, S.; Siribaddana, S., Drinking well wate Herbicides is associated with chronic kidney disease, in Padavi-Sripura, Sri Lanka. Environmenta	al 272 273
463-493.	, M. A.; Bonventre, J. V., Biomarkers of acute kidney injury. Annu Rev Pharmacol Toxicol 2008, 48	275
	6. S.; Ferguson, M. A.; Collings, F. B.; Sunderland, K.; Gioules, C.; Bradwin, G.; Matsouaka C., Urinary biomarkers for sensitive and specific detection of acute kidney injury in humans. Clinica (3), 200-208.	
injury: A systematic review. Kic	R.; Concato, J.; Parikh, C. R., Biomarkers for the diagnosis and risk stratification of acute kidney International 2008, 73 (9), 1008-1016.	280
	<, A.; Goldstein, S.; Kashani, K.; Macedo, E.; Murugan, R.; Bell, M.; Forni, L.; Guzzi, L.; Jo Legrand, M.; Mehta, R.; Murray, P. T.; Pickkers, P.; Plebani, M.; Prowle, J.; Ricci, Z.; Rim	
	A. D.; Kellum, J. A.; Ronco, C., Recommendations on Acute Kidney Injury Biomarkers From th re Consensus Conference: A Consensus Statement. JAMA Network Open 2020, 3 (10), e2019209	
12. Ichimura, T.; Hung, C. Comarker for nephrotoxicant-ind	C.; Yang, S. A.; Stevens, J. L.; Bonventre, J. V., Kidney injury molecule-1: a tissue and urinary bi luced renal injury. Am J Physiol Renal Physiol 2004, 286 (3), F552-63.	i- 286 287
Suppl 2008, 241, 78-83.	njury Molecule-1 (KIM-1): a specific and sensitive biomarker of kidney injury. Scand J Clin Lab Inves	289
2013, 394582.	asci, A. I., New Biomarkers for the Quick Detection of Acute Kidney Injury. ISRN Nephrology 2013	291
Molecule 1 for Acute Kidney Inj	W.; Zhang, Z.; Wang, C.; Qi, C.; Ni, Z.; Mou, S., Diagnostic Value of Urinary Kidney Injury jury: A Meta-Analysis. PLOS ONE 2014, 9 (1), e84131.	293
Orho-Melander, M., Plasma kid Dial Transplant 2020, 35 (2), 265		ol 295 296
madi, K. R.; Bakker, S. J. L.; B H. T.; Cotlarciuc, I.; Davey S	W.; Lord, G. M.; van der Harst, P.; Lawlor, D. A.; Sehmi, J. S.; Gale, D. P.; Wass, M. N.; Ah eckmann, J.; Bilo, H. J. G.; Bochud, M.; Brown, M. J.; Caulfield, M. J.; Connell, J. M. C.; Cook mith, G.; de Silva, R.; Deng, G.; Devuyst, O.; Dikkeschei, L. D.; Dimkovic, N.; Dockrell, M. Eggermann, T.; Farrall, M.; Ferrucci, L.; Floege, J.; Forouhi, N. G.; Gansevoort, R. T.; Han	k, 298 .; 299
X.; Hedblad, B.; Homan van P. E.; Kleefstra, N.; Lagou, V	der Heide, J. J.; Hepkema, B. G.; Hernandez-Fuentes, M.; Hypponen, E.; Johnson, T.; de Jong 7.; Lapsley, M.; Li, Y.; Loos, R. J. F.; Luan, J. a.; Luttropp, K.; Maréchal, C.; Melander, O. Parsa, A.; Peltonen, L.; Penninx, B. W.; Perucha, E.; Pouta, A.; Prokopenko, I.; Roderick, P. J.	g, 301 .; 302

Ruokonen, A.; Samani, N. J.; Sanna, S.; Schalling, M.; Schlessinger, D.; Schlieper, G.; Seelen, M. A. J.; Shuldiner, A. R.; 304 Sjögren, M.; Smit, J. H.; Snieder, H.; Soranzo, N.; Spector, T. D.; Stenvinkel, P.; Sternberg, M. J. E.; Swaminathan, R.; 305 Tanaka, T.; Ubink-Veltmaat, L. J.; Uda, M.; Vollenweider, P.; Wallace, C.; Waterworth, D.; Zerres, K.; Waeber, G.; Ware-306 ham, N. J.; Maxwell, P. H.; McCarthy, M. I.; Jarvelin, M.-R.; Mooser, V.; Abecasis, G. R.; Lightstone, L.; Scott, J.; Navis, G.; 307 Elliott, P.; Kooner, J. S., Genetic loci influencing kidney function and chronic kidney disease. Nat Genet 2010, 42 (5), 373-375. 308 18. Aguado-Fraile, E.; Ramos, E.; Conde, E.; Rodríguez, M.; Liaño, F.; García-Bermejo, M. L., MicroRNAs in the kidney: 309

novel biomarkers of acute kidney injury. Nefrología (English Edition) 2013, 33 (6), 826-834. 310 311

19. Holohan, K. N.; Lahiri, D. K.; Schneider, B. P.; Foroud, T.; Saykin, A. J., Functional microRNAs in Alzheimer's disease and cancer: differential regulation of common mechanisms and pathways. Front Genet 2013, 3, 323-323.

20. Wei, Q.; Mi, Q.-S.; Dong, Z., The regulation and function of micrornas in kidney diseases. IUBMB Life 2013, 65 (7), 602-614. 21. O'Brien, J.; Hayder, H.; Zayed, Y.; Peng, C., Overview of MicroRNA Biogenesis, Mechanisms of Actions, and Circulation. Front Endocrinol (Lausanne) 2018, 9, 402-402.

Condrat, C. E.; Thompson, D. C.; Barbu, M. G.; Bugnar, O. L.; Boboc, A.; Cretoiu, D.; Suciu, N.; Cretoiu, S. M.; 22. 316 Voinea, S. C., miRNAs as Biomarkers in Disease: Latest Findings Regarding Their Role in Diagnosis and Prognosis. Cells 2020, 9 (2), 317 276. 318

Weldon, B. A.; Shubin, S. P.; Smith, M. N.; Workman, T.; Artemenko, A.; Griffith, W. C.; Thompson, B.; Faustman, E. 23. M., Urinary microRNAs as potential biomarkers of pesticide exposure. Toxicology and applied pharmacology 2016, 312, 19-25.

Du, J.; Cao, X.; Zou, L.; Chen, Y.; Guo, J.; Chen, Z.; Hu, S.; Zheng, Z., MicroRNA-21 and Risk of Severe Acute Kidney 24. Injury and Poor Outcomes after Adult Cardiac Surgery. PLOS ONE 2013, 8 (5), e63390.

25. Gerlach, C. V.; Vaidya, V. S., MicroRNAs in injury and repair. Archives of Toxicology 2017, 91 (8), 2781-2797.

26. Chau, B. N.; Xin, C.; Hartner, J.; Ren, S.; Castano, A. P.; Linn, G.; Li, J.; Tran, P. T.; Kaimal, V.; Huang, X.; Chang, A. N.; Li, S.; Kalra, A.; Grafals, M.; Portilla, D.; MacKenna, D. A.; Orkin, S. H.; Duffield, J. S., MicroRNA-21 promotes fibrosis of the kidney by silencing metabolic pathways. Sci Transl Med 2012, 4 (121), 121ra18-121ra18.

Chung, A. C.; Dong, Y.; Yang, W.; Zhong, X.; Li, R.; Lan, H. Y., Smad7 suppresses renal fibrosis via altering expression 27. of TGF-β/Smad3-regulated microRNAs. Mol Ther 2013, 21 (2), 388-98.

28. Mohamed, F.; Endre, Z. H.; Buckley, N. A., Role of biomarkers of nephrotoxic acute kidney injury in deliberate poisoning and envenomation in less developed countries. British Journal of Clinical Pharmacology 2015, 80 (1), 3-19.

Jaikwang, P.; Junkuy, A.; Sapbamrer, R.; Seesen, M.; Khacha-ananda, S.; Mueangkhiao, P.; Wunnapuk, K., A Dilute-29. and-Shoot LC-MS/MS Method for Urinary Glyphosate and AMPA. Chromatographia 2020, 83 (3), 467-475.

Kao, H.-W.; Pan, C.-Y.; Lai, C.-H.; Wu, C.-W.; Fang, W.-L.; Huang, K.-H.; Lin, W.-C., Urine miR-21-5p as a potential 30. non-invasive biomarker for gastric cancer. Oncotarget 2017, 8 (34), 56389-56397.

Trasande, L.; Aldana, S. I.; Trachtman, H.; Kannan, K.; Morrison, D.; Christakis, D. A.; Whitlock, K.; Messito, M. J.; 31. 335 Gross, R. S.; Karthikraj, R.; Sathyanarayana, S., Glyphosate exposures and kidney injury biomarkers in infants and young children. 336 Environmental Pollution 2020, 256, 113334. 337

Abdul, K. S. M.; De Silva, P. M. C. S.; Ekanayake, E. M. D. V.; Thakshila, W. A. K. G.; Gunarathna, S. D.; Gunasekara, 32. 338 T. D. K. S. C.; Jayasinghe, S. S.; Asanthi, H. B.; Chandana, E. P. S.; Chaminda, G. G. T.; Siribaddana, S. H.; Jayasundara, N., 339 Occupational Paraquat and Glyphosate Exposure May Decline Renal Functions among Rural Farming Communities in Sri Lanka. 340 International Journal of Environmental Research and Public Health 2021, 18 (6), 3278. 341

De Silva, P. M. C. S.; Mohammed Abdul, K. S.; Eakanayake, E. M. D. V.; Jayasinghe, S. S.; Jayasumana, C.; Asanthi, H. 33. 342 B.; Perera, H. S. D.; Chaminda, G. G. T.; Chandana, E. P. S.; Siribaddana, S. H., Urinary Biomarkers KIM-1 and NGAL for Detec-343 tion of Chronic Kidney Disease of Uncertain Etiology (CKDu) among Agricultural Communities in Sri Lanka. PLOS Neglected Trop-344 ical Diseases 2016, 10 (9), e0004979. 345

34. Gil, H. W.; Yang, J. O.; Lee, E. Y.; Hong, S. Y., Clinical implication of urinary neutrophil gelatinase-associated lipocalin and kidney injury molecule-1 in patients with acute paraquat intoxication. Clin Toxicol (Phila) 2009, 47 (9), 870-5.

Kumarswamy, R.; Volkmann, I.; Thum, T., Regulation and function of miRNA-21 in health and disease. RNA Biol 2011, 8 (5), 35. 706-713.

Li, Y.-F.; Jing, Y.; Hao, J.; Frankfort, N. C.; Zhou, X.; Shen, B.; Liu, X.; Wang, L.; Li, R., MicroRNA-21 in the patho-36. genesis of acute kidney injury. Protein & Cell 2013, 4 (11), 813-819.

Pavkovic, M.; Robinson-Cohen, C.; Chua, A. S.; Nicoara, O.; Cárdenas-González, M.; Bijol, V.; Ramachandran, K.; 37. Hampson, L.; Pirmohamed, M.; Antoine, D. J.; Frendl, G.; Himmelfarb, J.; Waikar, S. S.; Vaidya, V. S., Detection of Drug-Induced Acute Kidney Injury in Humans Using Urinary KIM-1, miR-21, -200c, and -423. Toxicol Sci 2016, 152 (1), 205-213.

38. Zhao, H.-W.; Liu, H.; Liu, L.-Y.; Liu, Z.; Dong, X.-S., Analysis of microRNA expression profiling during paraquat-induced injury of murine lung alveolar epithelial cells. The Journal of Toxicological Sciences 2020, 45 (8), 423-434.

Chen, C.; Lu, C.; Qian, Y.; Li, H.; Tan, Y.; Cai, L.; Weng, H., Urinary miR-21 as a potential biomarker of hypertensive 39. kidney injury and fibrosis. Scientific Reports 2017, 7 (1), 17737.

Garrigou, A.; Laurent, C.; Berthet, A.; Colosio, C.; Jas, N.; Daubas-Letourneux, V.; Jackson Filho, J. M.; Jouzel, J. N.; 40. 359 Samuel, O.; Baldi, I.; Lebailly, P.; Galey, L.; Goutille, F.; Judon, N., Critical review of the role of PPE in the prevention of risks 360 related to agricultural pesticide use. Safety Science 2020, 123, 104527. 361

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